

Bovine Interleukin -1 Beta (bIL-1 β) AlphaLISA Immunoassay Kit

Product number: AL539HV/C/F

Research Use Only. Not for use in diagnostic procedures.

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Product Information

- Application:** This kit is designed for the quantitative determination of bovine IL-1 beta (bIL-1 β) buffer and cell culture media using a homogeneous AlphaLISA assay (no wash steps). The assay shows negligible cross-reactivity with other bovine cytokines.
- Sensitivity:** Lower Detection Limit (LDL): 20 \pm 15 pg/mL
Lower Limit of Quantification (LLOQ): 67 \pm 45 pg/mL
EC₅₀: 37 \pm 12 ng/mL
- Dynamic range:** 1 - 300000 pg/mL (Figure 1).

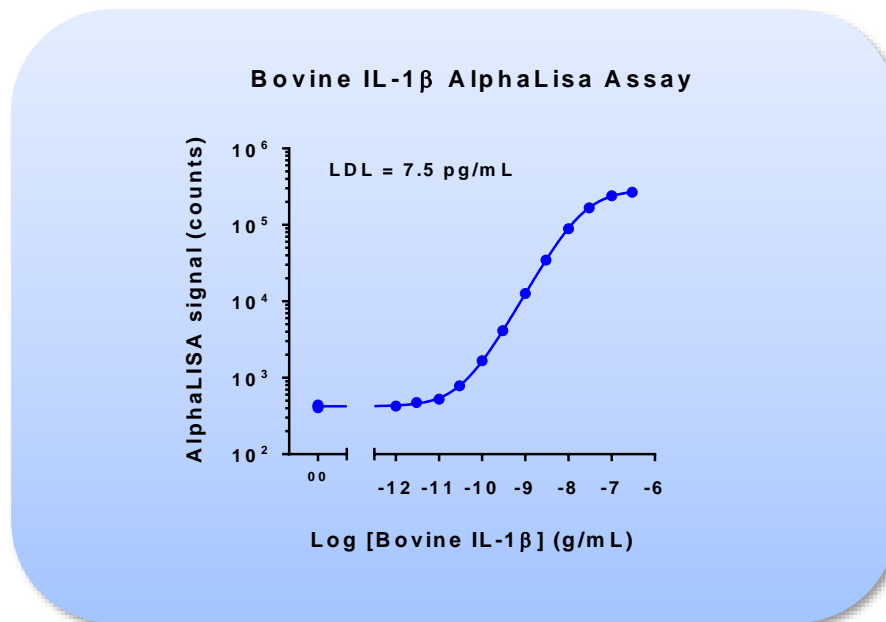


Figure 1. Typical sensitivity curves in AlphaLISA Immunoassay Buffer. The data was generated using a white Optiplate™-384 microplate and the EnVision® Multilabel Plate Reader with Alpha option 2102.

- Storage:** Store kit in the dark at +4°C. Store reconstituted analyte at -20°C.
- Stability:** This kit is stable for at least 6 months from the manufacturing date when stored in its original packaging and the recommended storage conditions. Note: Once reconstituted, the bovine IL-1 β analyte is stable for at least 18 months when stored at -20°C.

Quality Control

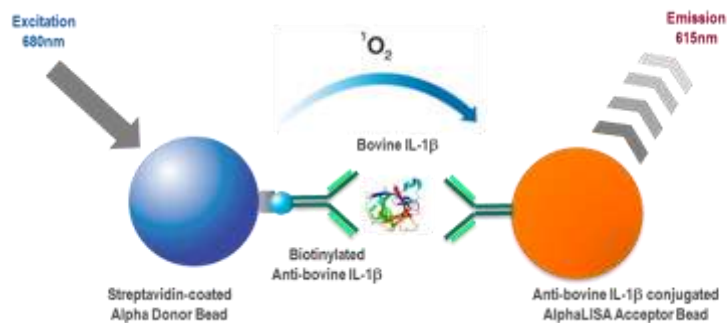
Lot to lot consistency is confirmed in an AlphaLISA assay. Maximum and minimum signals, EC₅₀ and LDL were measured on the EnVision Multilabel Plate Reader with Alpha option using the protocol described in this technical data sheet. We certify that these results meet our quality release criteria. Maximum counts may vary between bead lots and the instrument used, with no impact on LDL measurement.

Analyte of Interest

IL-1, a proinflammatory cytokine, is produced by immune cells following bacterial, viral, fungal, and parasitic infections. It is produced as two forms, IL-1 α and IL-1 β . IL-1 β mediates both local and systemic inflammatory responses and up-regulation of bovine IL-1 β in serum and milk is associated bovine with mastitis diseases and to vaccination.

Description of the AlphaLISA Assay

AlphaLISA technology allows the detection of molecules of interest in buffer, cell culture media, serum and plasma in a highly sensitive, quantitative, reproducible and user-friendly mode. In an AlphaLISA assay, a Biotinylated Anti-Analyte Antibody binds to the Streptavidin-coated Alpha Donor beads, while another Anti-Analyte Antibody is conjugated to AlphaLISA Acceptor beads. In the presence of the analyte, the beads come into close proximity. The excitation of the Donor beads provokes the release of singlet oxygen molecules that triggers a cascade of energy transfer in the Acceptor beads, resulting in a sharp peak of light emission at 615 nm (Figure 2).



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Figure 2. AlphaLISA Assay principle.

Precautions

- The Alpha Donor beads are light-sensitive. All the other assay reagents can be used under normal light conditions. All Alpha assays using the Donor beads should be performed under subdued laboratory lighting (< 100 lux). Green filters (LEE 090 filters (preferred) or Roscolux filters #389 from Rosco) can be applied to light fixtures.
- All blood components and biological materials should be handled as potentially hazardous. The analyte included in this kit is from a bovine source.
- Some analytes are present in saliva. Take precautionary measures to avoid contamination of the reagent solutions.
- The Biotinylated Anti-Analyte Antibody contains sodium azide. Contact with skin or inhalation should be avoided.

Kit Content: Reagents and Materials

Kit components	AL539HV (100 assay points ^{***})	AL539C (500 assay points ^{***})	AL539F (5000 assay points ^{***})
AlphaLISA Anti- bIL-1 β Acceptor beads stored in PBS, 0.05% Proclin-300, pH 7.2	20 μ L @ 5 mg/mL (1 brown tube, <u>white</u> cap)	50 μ L @ 5 mg/mL (1 brown tube, <u>white</u> cap)	500 μ L @ 5 mg/mL (1 brown tube, <u>white</u> cap)
Streptavidin (SA)-coated Donor beads stored in 25 mM HEPES, 100 mM NaCl, 0.05% Proclin-300, pH 7.4	100 μ L @ 5 mg/mL (1 brown tube, <u>black</u> cap)	200 μ L @ 5 mg/mL (1 brown tube, <u>black</u> cap)	2 mL @ 5 mg/mL (1 brown tubes, <u>black</u> caps)
Biotinylated Antibody Anti- bIL-1 β stored in PBS, 0.1% Tween-20, 0.05% NaN ₃ , pH 7.4	20 μ L @ 500 nM (1 tube, <u>black</u> cap)	50 μ L @ 500 nM (1 tube, <u>black</u> cap)	500 μ L @ 500 nM (1 tube, <u>black</u> cap)
AlphaLISA bIL-1 β (0.3 μ g), lyophilized analyte *	1 tube, <u>clear</u> cap	1 tube, <u>clear</u> cap	1 tube, <u>clear</u> cap
AlphaLISA Immunoassay Buffer (10X) **	2mL, 1 small bottle	10 mL, 1 small bottle	100 mL, 1 large bottle

* Reconstitute bIL-1 β in 100 μ L Milli-Q[®] grade H₂O. The reconstituted analyte should be used within 60 minutes or aliquoted into screw-capped polypropylene vials and stored at -20°C for further experiments. Avoid multiple freeze-thaw cycles. It has been demonstrated that reconstituted bIL-1 β is stable for at least 18 months at -20°C. One vial contains an amount of bIL-1 β sufficient for performing 10 standard curves. Additional vials can be ordered separately (cat # AL539S).

** Extra buffer can be ordered separately (cat # AL000C: 10 mL, cat # AL000F: 100 mL).

*** The number of assay points is based on an assay volume of 100 μ L in 96-well plates (AL522HV) or 50 μ L in 96- or 384-well assay plates using the kit components at the recommended concentrations.

Sodium azide should **not** be added to the stock reagents. High concentrations of sodium azide (> 0.001 % final in the assay) might decrease the AlphaLISA signal. Note that sodium azide from the Biotinylated Antibody stock solution will not interfere with the AlphaLISA signal (0.0001% final in the assay).

Specific additional required reagents and materials:

The following materials are recommended:

Item	Suggested source	Catalog #
TopSeal™-A Adhesive Sealing Film	PerkinElmer Inc.	6050195
EnVision®-Alpha Reader	PerkinElmer Inc.	-

Recommendations

General recommendations:

- The volume indicated on each tube is guaranteed for single pipetting. Multiple pipetting of the reagents may reduce the theoretical amount left in the tube. To minimize loss when pipetting beads, it is preferable not to pre-wet the tip.
- Centrifuge all tubes (including lyophilized analyte) before use to improve recovery of content (2000g, 10-15 sec). Re-suspend all reagents by vortexing before use.
- Use Milli-Q® grade H₂O (18 MΩ•cm) to dilute 10X AlphaLISA Immunoassay Buffer to reconstitute the lyophilized analyte.
- When diluting the standard or samples, change tips between each standard or sample dilution. When loading reagents in the assay microplate, change tips between each standard or sample addition and after each set of reagents.
- When reagents are added to the microplate, make sure the liquids are at the bottom of the well.
- Small volumes may be prone to evaporation. It is recommended to cover microplates with TopSeal-A Adhesive Sealing Films to reduce evaporation during incubation. Microplates can be read with the TopSeal-A Film.
- The AlphaLISA signal is detected with an EnVision Multilabel Reader equipped with the Alpha option using the AlphaScreen standard settings (e.g. Total Measurement Time: 550 ms, Laser 680 nm Excitation Time: 180 ms, Mirror: D640as, Emission Filter: M570w, Center Wavelength 570 nm, Bandwidth 100 nm, Transmittance 75%).
- AlphaLISA signal will vary with temperature and incubation time. For consistent results, identical incubation times and temperature should be used for each plate.
- The standard curves shown in this technical data sheet are provided for information only. A standard curve must be generated for each experiment.

Assay Procedure

IMPORTANT: PLEASE READ THE RECOMMENDATIONS BELOW BEFORE USE

- The protocol described below is an example for generating one standard curve in a 50 µL final assay volume (48 wells, triplicate determinations). The protocols also include testing samples in 354 wells. If a different amount of samples are tested, the volumes of all reagents have to be adjusted accordingly, as shown in the table below. These calculations do not include excess reagent to account for losses during transfer of solutions or dead volumes.
- The standard dilution protocol is provided for information only. As needed, the number of replicates or the range of concentrations covered can be modified.
- Use of four background points in triplicate (12 wells) is recommended when LDL/LLOQ is calculated. One background point in triplicate (3 wells) can be used when LDL/LLOQ is not calculated.

Format	# of data points	Volume				Plate recommendation
		Final	Sample	AlphaLISAbeads / Biotin Antibody MIX	SA-Donor beads	
AL539HV	100	100 µL	10 µL	40 µL	50 µL	White OptiPlate-96 (cat # 6005290) White ½ AreaPlate-96 (cat # 6005560)
AL539C	250	100 µL	10 µL	40 µL	50 µL	White OptiPlate-96 (cat # 6005290) White ½ AreaPlate-96 (cat # 6005560)
	500	50 µL	5 µL	20 µL	25 µL	White ½ AreaPlate-96 (cat # 6005560) White OptiPlate-384 (cat # 6007290) Light gray AlphaPlate™-384 (cat # 6005350)
	1 250	20 µL	2 µL	8 µL	10 µL	Light gray AlphaPlate-384 (cat # 6005350) ProxiPlate™-384 Plus (cat # 6008280) White OptiPlate-384 (cat # 6007290)
	2 500	10 µL	1 µL	4 µL	5 µL	Light gray AlphaPlate-1536 (cat # 6004350)
AL539F	5 000	50 µL	5 µL	20 µL	25 µL	White ½ AreaPlate-96 (cat # 6005560) White OptiPlate-384 (cat # 6007290) Light gray AlphaPlate-384 (cat # 6005350)
	12 500	20 µL	2 µL	8 µL	10 µL	Light gray AlphaPlate-384 (cat # 6005350) ProxiPlate-384 Plus (cat # 6008280) White OptiPlate-384 (cat # 6007290)
	25 000	10 µL	1 µL	4 µL	5 µL	Light gray AlphaPlate-1536 (cat # 6004350)

The 3-step high sensitivity protocol described below is for 500 assay points including one standard curve (48 wells) and samples (452 wells).

1) Preparation of 1X AlphaLISA Immunoassay Buffer :

- Add 10 mL of 10X AlphaLISA Immunoassay Buffer to 90 mL H₂O.

2) Preparation of bIL-1 β analyte standard dilutions:

- bIL-1 β analyte is provided at 0.3 μ g in lyophilized form. Reconstitute with 100 μ L MiliQ H₂O to create a 3 μ g/mL solution. The first point of the curve is 0.3 μ g/mL so a 10 fold dilution is required. Prepare standard dilutions as follows (change tip between each standard dilution):

Tube	Vol. of bIL-1 β (μ L)	Vol. of diluent (μ L) *	[bIL-1 β] in standard curve	
			(g/mL in 5 μ L)	(pg/mL in 5 μ L)
A	10 μ L of provided bIL-1 β	90	3.00E-07	300000
B	60 μ L of tube A	120	1.00E-07	100000
C	60 μ L of tube B	140	3.00E-08	30000
D	60 μ L of tube C	120	1.00E-08	10000
E	60 μ L of tube D	140	3.00E-09	3000
F	60 μ L of tube E	120	1.00E-09	1000
G	60 μ L of tube F	140	3.00E-10	300
H	60 μ L of tube G	120	1.00E-10	100
I	60 μ L of tube H	140	3.00E-11	30
J	60 μ L of tube I	120	1.00E-11	10
K	60 μ L of tube J	140	3.00E-12	3
L	60 μ L of tube K	120	1.00E-12	1
M ** (background)	0	100	0	0
N ** (background)	0	100	0	0
O ** (background)	0	100	0	0
P ** (background)	0	100	0	0

* Dilute standards in diluent (e.g. 1X AlphaLISA Immunoassay Buffer).

At low concentrations of analyte, a significant amount of analyte can bind to the vial. Therefore, load the analyte standard dilutions in the assay microplate within 60 minutes of preparation.

** Four background points in triplicate (12 wells) are used when LDL is calculated. If LDL does not need to be calculated, one background point in triplicate can be used (3 wells).

3) Preparation of 5X AlphaLISA anti-bIL-1 β Acceptor beads (50 μ g /mL)

Add 60 μ L of 5 mg/mL AlphaLISA anti- bIL-1 β acceptor beads to 5940 μ L of AlphaLISA Immunoassay Buffer .

Prepare just before use.

4) Preparation of 5X biotinylated anti- bIL-1 β antibody (5 nM):

Add 60 μ L of 500nM biotinylated anti- bIL-1 β antibody to 5940 μ L of 1X AlphaLISA Bovine Immunoassay Buffer.

Prepare just before use.

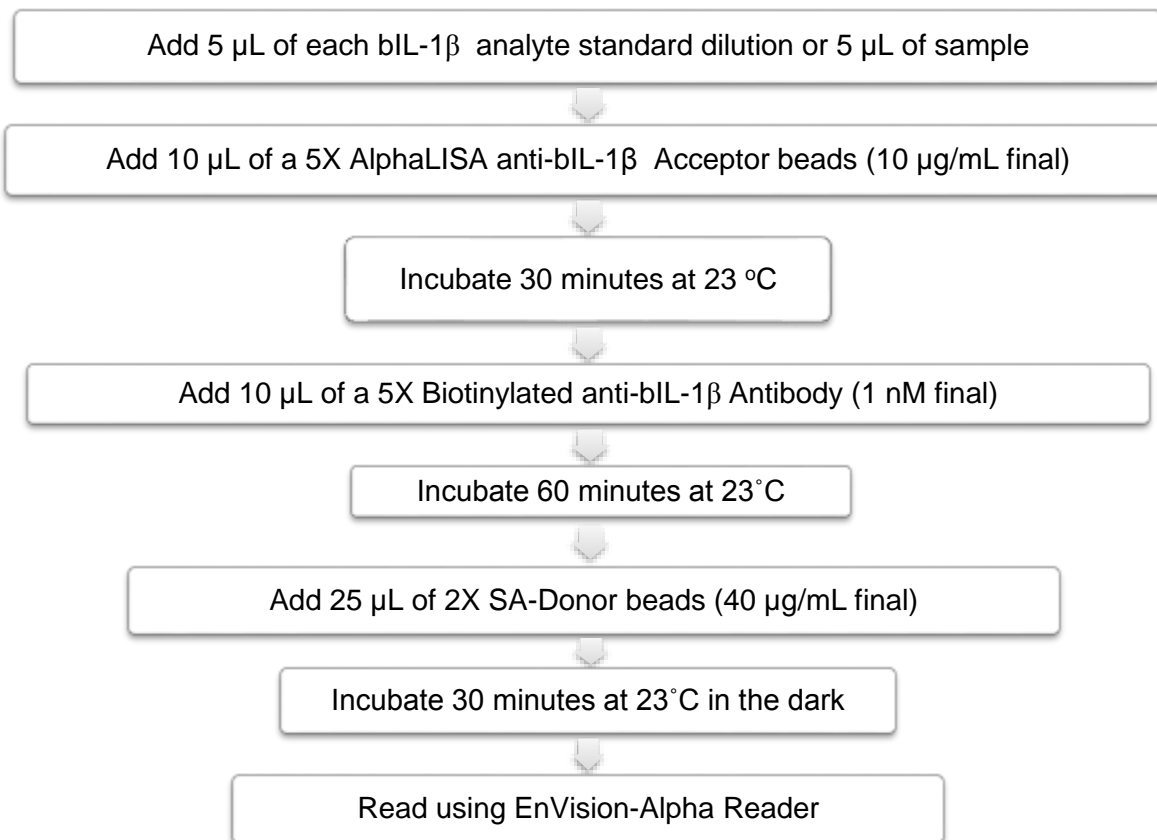
5) Preparation of 2X Streptavidin (SA) Donor beads (80 μ g/mL):

Keep the beads under subdued laboratory lighting.

Add 200 μ L of 5 mg/mL SA-Donor beads to 12 300 μ L of 1X AlphaLISA Immunoassay Buffer.

Prepare just before use.

6) In a 96- or 384-well microplate:



Data Analysis

- Calculate the average count value for the background wells.
- Generate a standard curve by plotting the AlphaLISA counts versus the concentration of analyte. A log scale can be used for either or both axes. No additional data transformation is required.
- Analyze data according to a nonlinear regression using the 4-parameter logistic equation (sigmoidal dose-response curve with variable slope) and a $1/Y^2$ data weighting (the values at maximal concentrations of analyte after the hook point should be removed for correct analysis).
- The LDL is calculated by interpolating the average background counts (12 wells without analyte) + 3 x standard deviation value (average background counts + (3xSD)) on the standard curve.
- The LLOQ as measured here is calculated by interpolating the average background counts (12 wells without analyte) + 10 x standard deviation value (average background counts + (10xSD)) on the standard curve. Alternatively, the true LLOQ can be determined by spiking known concentrations of analyte in the matrix and measuring the percent recovery, and then determining the minimal amount of spiked analyte that can be quantified within a given limit (usually +/- 20% or 30% of the real concentration).
- Read from the standard curve the concentration of analyte contained in the samples.
- If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

Assay Performance Characteristics

AlphaLISA assay performance described below was determined using the 3 step protocol.

- Assay Sensitivity:

The LDL and LLOQ were calculated as described above. The values correspond to the lowest concentration of analyte that can be detected in a volume of 5 μ L using the recommended assay conditions.

LDL (pg/mL)	Buffer/Media	# of experiments
15	AlphaLISA Immunoassay Buffer	25
24	DMEM+ 10% FBS	7
23	RPMI + 10% FBS	7

* Note that LDL/ LLOQ can be decreased (i.e. sensitivity increased) by increasing the volume of analyte in the assay (e.g. use 10 μ L of analyte in a final assay volume of 50 μ L).

- Assay Precision:

The following assay precision data were calculated from the three independent assays using two different kit lots. In each lot, the analytes were prepared in AlphaLISA Immunoassay Buffer (IAB), DMEM, or RPMI. Each assay consisted of one standard curve comprising 12 data points (each in triplicate) and 12 background wells (no analytes). The assays were performed in 384-well format using AlphaLISA Immunoassay Buffer.

- Intra-assay precision:

The intra-assay precision was determined using a total of 16 independent determinations in triplicate. Shown are CV%.

bIL-1 β	IAB	DMEM	RPMI
CV%	5.1%	3.5%	4.9%

- Inter-assay precision:

The inter-assay precision was determined using a total of 3 independent determinations with 9 measurements for 3 ng/mL sample. Shown are CV%.

bIL-1 β (3 ng/ml)	IAB	DMEM	RPMI
CV%	13.2%	9.2%	10.9%

- Spike Recovery:

Three known concentrations of analyte were spiked in IAB, or in cell culture media containing 10% FBS. All samples, including non-spiked Immunoassay Buffers and culture media were measured in the assay. The average recovery from three independent measurements is reported.

Spiked bIL-1 β (ng/mL)	% Recovery		
	IAB	DMEM*	RPMI*
10	99	71 (98)	62 (109)
3	85	65 (105)	71 (98)
0.3	110	52 (112)	74 (110)

Note: * the numbers in parenthesis are the % recovery using the standards made in cell culture media.

- Specificity:

Cross-reactivity of the AlphaLISA bIL-1 β Kit was tested using the following proteins at 100 ng/mL in IAB. Reactivity to bIL-1 β is 100%.

Protein	% Cross-reactivity
Bovine IL-1 α	0.02
Bovine IL-2	0.04
Bovine IL-6	0.02
Bovine IFN γ	0.04
Bovine TNF β	0.02
Swine IL-1 β	0.03
Equine IL-1 β	0.03
Ovine IL-1 β	16.88
Human IL-1 β	0.03

Troubleshooting Guide

You will find detailed recommendations for common situations you might encounter with your AlphaLISA Assay kit at:

http://www.perkinelmer.com/in/resources/technicalresources/applicationsupportknowledgebase/alphalisa-phascreen-no-washassays/alpha_troubleshoot.xhtml

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