

Development of a high throughput SPA assay using a cloned kappa opioid receptor.

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Introduction

Opioid receptors have been shown to be capable of producing strong analgesic effects when activated by opiates^{1,2} and their potential role in pain relief is being investigated. Opioid receptor classes have been identified by the pharmacology of a range of selective ligands, these have been named mu (μ), delta (δ), and kappa (κ).

Amersham Biosciences has developed a scintillation proximity assay (SPA) for high throughput screening using a κ -opioid receptor membrane prepared from cloned HEK-293 cells (SignalScreen, Amersham Biosciences UK Ltd. 6110558 200U). A range of ligands known to have a high affinity for the κ -opioid receptor was used to generate IC₅₀ values in this assay.

Methods

1mg WGA coated polyvinyl toluene (PVT) SPA bead, 15 μ g membrane and [³H]diprenorphine (DPN) (concentration as shown below), were incubated for 18 hours at room temperature. Non specific binding (NSB) was determined in the presence of 50 μ M unlabelled naloxone. All assays were performed in a total volume of 200 μ l assay buffer: 50mM tris-HCl, pH7.4.

For competition binding curves, a range of concentrations of β -endorphin was prepared using buffer. Naltrexone and U50-488 were prepared using DMSO; final concentration of DMSO in well was 2.5%.

Results

Saturation binding experiments were carried out to determine K_d and Bmax. A range of concentrations of [³H]DPN was prepared in assay buffer to give final concentrations as shown in figure 1.

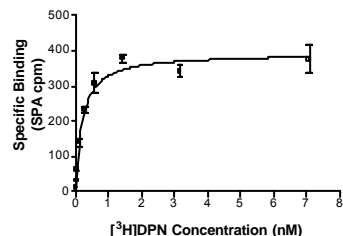


Figure 1. Results of saturation binding experiment. Values are means \pm SD (n=3)

Bmax and K_d were determined to be 1.99pmol/mg and 0.22nM respectively, in the SPA assay (experiments n=3), and were similar to those obtained in our experiments in filter binding assays: Bmax 1.75 pmol/mg, K_d 0.21nM (data not shown).

For high throughput screening purposes, a 96-well assay plate was used and the volume was therefore restricted to a maximum of 200 μ l. At [³H]DPN concentration 0.2nM (K_d) the assay was shown to be in ligand depletion (>25% of added ligand was bound). In order to avoid this, and therefore accurately determine IC₅₀ values, the ligand concentration was increased to 1.8nM. Under these conditions ligand depletion was <10%.

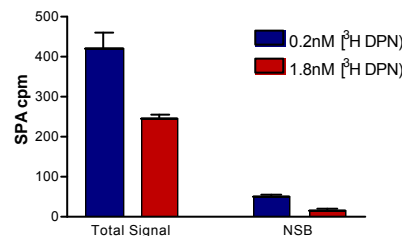


Figure 2. Total signal and NSB at [³H]DPN concentrations of 0.2nM and 1.8nM

Increasing [³H]DPN concentration from 0.2nM to 1.8nM reduced the assay signal window (difference between total signal and NSB) from 17:1 to 9:1. The Z' factor⁷ was calculated at the increased ligand concentration (1.8nM), this demonstrated that the assay was a good screening assay with a Z' of 0.76 (figure 3).

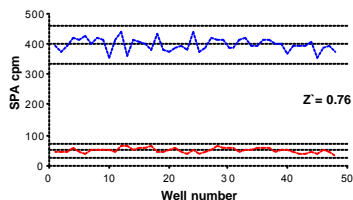


Figure 3. Z' factor analysis of SPA assay, total binding (—) NSB (···). Solid lines indicate mean of 48 observations, dotted lines indicate mean \pm 3 x SD.

The assay was also tested for tolerance to DMSO, a range of concentrations was included from 0-9% (v/v) (figure 4). The assay was tolerant to a DMSO concentration of at least 4.5% (v/v).

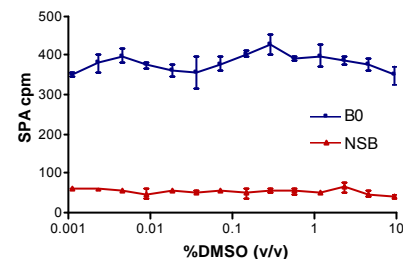


Figure 4. Total assay signal obtained at range of concentrations of DMSO from 0 - 9% (v/v). Values are means \pm SD (n=3).

Competition binding studies (figure 5) were performed using a range of ligands that have been shown to be selective for the κ -opioid receptor.^(3, 4, 5)

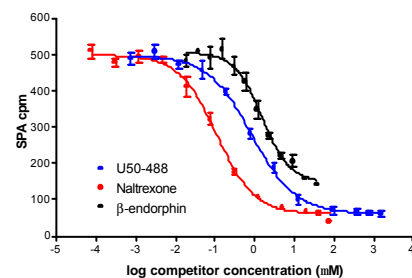


Figure 5. Binding of [³H]DPN to κ -opioid receptor captured with WGA coated PVT SPA beads. Competition with β -endorphin (·), naltrexone (·) and U50-488 (·). Values are means \pm SEM (replicates n=3). IC₅₀ values are shown in table 2.

Ligand	IC ₅₀ (μ M)	95% Confidence Intervals	Selective for subtype
U50-488	0.669	0.557-0.809	? ₁ & ? ₃
Naltrexone	0.082	0.065-0.104	? ₁
β -Endorphin	0.907	0.680-1.219	? ₂

Table 1. IC₅₀ values obtained for each subtype specific ligand tested (experiments n=3).

CONCLUSIONS

- A high throughput-screening assay to identify compounds that bind to a cloned κ -opioid receptor has been developed.
- Tolerance to DMSO concentration up to at least 4.5% (v/v) has been demonstrated.
- Competition binding studies indicate that the ? subtype is predominantly expressed in this membrane preparation. This confirms previous studies that cloned κ -opioid receptors exhibit typical pharmacological K₁ profile.^(5, 6)

References

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